Please read these and keep them in mind as you do your experiments!

* Reduce the potential to contaminate your surroundings by working in a clean, uncluttered space.
* Be organized and read through each experiment thoroughly before starting. Gather and prepare all necessary reagents and instruments before beginning the experiments.
* Immediately cover plates after pouring media into them. When opening a plate to either inoculate it or to transfer bacteria from it, use the lid as a shield to limit contamination from airborne sources.
* When using tubes containing media or other liquids, hold the tube at an angle and immediately recap to limit contamination from airborne sources.
* Always use a new, clean, sterile, disposable loop or transfer pipette when transferring multiple samples or colonies of bacteria, unless otherwise directed in the experiment instructions.
* Store prepared, solidified agar plates (not yet inoculated) upside-down so that any condensation that forms on them will not drop onto the media surface.
* Disinfect all cultures at the end of every experiment (unless otherwise instructed) by adding a 10% bleach solution to the culture plates and letting them sit at room temperature for 20 minutes. Household bleach contains 5.25% sodium hypochlorite, and is corrosive so surfaces should be wiped with water after using bleach. A 10% solution of household bleach will kill many organisms immediately, but may take up to 20 minutes for full sterilization.
* After flooding the plates, pour the bleach down a drain, seal the plates with Parafilm™, and dispose of them in the trash.
* Be sure to clean and disinfect your work area with disinfecting wipes or 70% ethanol after each experiment.